Springfield Technical Community College

ACADEMIC AFFAIRS

Course Number:	CLLS 316	Department:	Clinical Laboratory Sciences		
Course Title:	Immunology and Immunohematology	Semester:	Spring	Year:	2014

Objectives/Competencies

Competencies	Course Objective		
Students will possess an understanding of the course expectations and scope	 Review all CLLS affective behaviors required to be a laboratory professional Discuss student responsibilities as they relate to an active learning classroom Review semester calendar and due dates 		
2. Students will apply all safety protocols, universal precautions when practicing laboratory skills. (Re: CLLS-103)	Practice all safety protocols and apply universal precautions when performing laboratory skills in the student laboratory.		
3. Students will possess an understanding of basic Immunology terminology.	 Describe the difference between innate and adaptive immunity. Describe the difference between cellular and humoral immunity. Define what makes an antigen. 		
4. Students will list the characteristics of antigens and antibodies.	 Describe the structure of a typical antibody List the characterists of the 5 immunoglobulin types found in humans. 		
5. Students will describe the difference between types of immunity.	 Describe innate v adaptive immunity, primary v secondary immunity and cell-mediated v humoral immunity Describe passive immunity 		
6. Students will discuss the use and makeup of common vaccinations	Discuss the role of adjuvants.		

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7. Students will understand the types of cells involved in the process of Innate Immunity	 List the types of granulocytes and mononuclear cells involved in innate immunity Describe the function of each cell and instances in which they are elevated. 		
8. Students will describe the other body functions involved in the innate immune response including Phagocytosis and Inflammation.	 List the steps in the process of phagocytosis. Explain the importance of phagocytosis in both natural and acquired immunity. Describe the process of inflammation. 		
9. Students will identify the types of cells involved in the Processes of Adaptive Immunity.	 List the acute phase reactants involved in innate immunity Describe the maturation process of T cells and B cells. Describe the role of B cells and T cells in immunity. 		
10. Students will describe the MHC – Major Histocompatibility Complex and its function.	 Define the MHC and discuss their function in regards to T cell activation. Discuss the differences between class I and class II molecules and their roles in adaptive immunity. 		
 11. Students will list the Soluble Mediators and describe their function within the immune response: Complement, Cytokines, Interleukins, and Interferons The Complement System 	 Identify the components of the complement system Discuss the complement activation pathways Describe the effects of increased or decreased complement on the immune system 		
10. Students will list the types of methodology used in Serology Testing – Laboratory techniques	 Define avidity and affinity List and describe methods in which precipitation reactions may be measured Describe the steps involved in the agglutination process Review and describe the types of labeled immunoassays and how they are used 		

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 11. Students will describe the body's Immune Response to Infectious diseases such as: Syphilis, Hepatitis, HIV, Infectious Mononucleosis, Streptococcal infections 	 Discuss automated methods to measure antigen-antibody reactions. Describe important characteristics of infectious disease Describe the body's response to infectious disease Differentiate the response of the immune system to intracellular and extracellular organisms List specific examples of bacterial and viral infections and describe the body's immune response 		
 12. Students will understand the mechanisms that lead to Immunologically related disorders such as: Hypersensitivity Immunoproliferative disorders Autoimmune Disease – Lupus, Rheumatoid arthritis, 	 Describe the general characteristics of autoimmune disorders Describe the role of the immune system in autoimmune disorders Describe the types and mechanisms of hypersensitivity Describe the specific laboratory tests to diagnose SLE and RA Describe the various fluorescent ANA patterns in the diagnosis of SL. 		
14. Students will review and apply medical terminology and general knowledge, principles of immunology concepts as they apply to the immunohematology laboratory	 Describe the Ag/Ab reaction and how we visualize it in the laboratory Give examples of agglutination, hemolysis, and neutralization reactions and describe the appearance of a positive and negative result List at least 3 factors that affect agglutination Define AHG and list its applications in the blood bank laboratory Discuss and compare the characteristics of IgG and IgM antibodies and determine which class will; React at room temp or 37 degrees C Cross the placenta Activate complement Be part of the primary or secondary immune response and in what amounts 		

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15. Specimen Collection	 List the types of specimens received in the blood bank laboratory and select the most appropriate blood tube to be drawn for each test Discuss the types of additives and anticoagulants in phlebotomy tubes and how they may interfere with blood bank testing Discuss the importance of specimen identification and some of the specific procedures employed in the blood bank lab to keep patients safe Assess the sample quality in regards to hemolysis and date/time of draw for specific testing 		
16. Students will review and apply the principles of genetics and inheritance.	 Prepare an inheritance chart and label the meaning of all symbols State the definition of phenotype vs. genotype Discuss the concepts of recessive and dominant traits in regards to inheritance Interpret a sex-linked inheritance chart and explain whether sons or daughters will inherit the trait. Discuss genes and alleles and how they relate to inheritance Determine genotypes for offspring using Punnett squares 		
 17. Students will possess a working knowledge of the ABO system and be able to perform forward and reverse ABO typing in the lab. ABO antigens ABO antibodies Secretor status Subgroups Testing reagents and procedures Troubleshooting discrepancies 	 List the possible genotypes for each ABO phenotype State which chromosome(s) ABH genes are located on Determine which sugars on the RBC are responsible for gene expression Describe how ABO antigen type determines the ABO antibody in the serum. List the reagents and patient cells/serum that are added to each tube in the forward and reverse ABO typing procedure List common reasons for discrepancies in ABO testing and 		

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19. Discuss the RBC antigen system that determines Rh status and perform Rh testing in the laboratory	determine which type of blood should be given if a discrepancy is found. Recognize the presence or absence of agglutination while performing the tilt tube method with an agglutination reader Explain the secretor test and interpret results Discuss the relation of the D antigen to Rh status Describe the different types of nomenclature and be able to translate between the three methods Categorize Rh antibodies as IgG or IgM immunoglobulins and discuss how they are formed		
20.Students will discuss the other common blood groups including; Kell, Duffy, Kidd, MNS, Lewis, Ii, and Diego	 discuss how they are formed. Perform Rh testing in the laboratory using the tilt tube method Summarize the procedure for Rh testing including Weak D testing List all reagents used in Rh testing and describe control reagent List the other RBC antigen groups including Kell, Duffy, Kidd, Lewis, Ii, MNS, and Diego 		
	 Using class notes, complete a chart that describes the antigens, antibodies, inheritance, fetal development and clinical significance. Describe what is unique to each blood group. Define dosage and give an example of its application to antibody testing. 		
21. Students will describe and perform antibody testing methods	 Explain the difference between the IAT and DAT List uses for each test Define zeta potential and how it is related to agglutination Determine the blood type and number of cells used for panels List the reagents used in the antibody screen and their uses Observe/explain gel testing techniques 		
	Understand the concept of heterozygous and homozygous cells		

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22. Students will describe and perform antibody identification techniques	 Understand how clinical significance is related to type of immunoglobulin and reaction temperature Practice and demonstrate the Elimination method, inclusion method, and rule of threes. Define other techniques such as: enzyme treatment, elution, adsorption, and absorption
23. Students will determine compatibility between recipient serum and donor red blood cells	 Understand the importance of patient identification in blood bank testing Explain the different types of crossmatches and when they are used: Immediate spin, Abbreviated, Antiglobulin, and Computer List, in order of preference, which type RBC unit would be used for each patient
24. Students will identify and give examples of various blood components and appropriate patient uses for such products	 List the type of blood products available and describe the use. Determine what type of blood is needed for each type of transfusion in terms of antigen type, antibody type, and preparation of cells (irradiation, CMV status).
25. Students will describe methods used for blood product processing and storage.	 Explain how each of the following blood products are prepared; Whole blood, Packed Red Blood Cells, FFP, platelet concentrates, cryoprecipitate List the optimal storage temperatures and maximum shelf life for each product Determine the blood type of universal donor and universal recipient for whole blood, PRBC, and FFP Describe the procedures that are available to remove excess WBC's from blood products

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26. Students will discuss the possible adverse effects of blood product transfusions	 List the possible adverse affects of transfusions and explain how they may occur List and define the different types of transfusion reactions including; IHTR, febrile, urticaria, bacterial, and DHTR List and explain the steps in a post-transfusion reaction workup Determine the type of post-transfusion specimen(s) needed for a reaction workup List and define the different types of transfusion including: autologous, intrauterine, and exchange Give an example in which you would use each type of transfusion 		
27. Students will identify donor selection protocols and procedures	 Demonstrate patient identification procedure in the blood bank Demonstrate specimen labeling criteria in the blood bank Recognize normal ranges for pre-collection testing results List the serology tests performed on blood products List the components of the donor screening process List the conditions that require permanent or temporary deferral of a blood donor Determine the volume of blood drawn for a unit of blood Give an example of confidentiality issues that may occur with blood donation, and describe how the issue may be resolved. 		
28. Students will discuss the causes and clinical manifestations of hemolytic disease of the newborn.	 Describe how HDN occurs List the antibodies most commonly responsible for the disease Describe common testing methods form fetal-maternal bleeding including the rosette test and Kleihauer-Betke Define Rhogam and explain when it is given to mothers Relate dosage of Rhogam to volume of FMH 		

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30. Students will briefly describe the HLA system	 Describe how the HLA typing is determined List situations in which HLA typing is used 		
31. Students will adhere to all affective behavioral objectives	 1. Safety a. Comply with all established laboratory safety regulations including: i. Standard precautions including PPE use and handwashing. ii. Practice proper handling and disposal of biohazardous materials. iii. Proper handling and disposal of sharps. iv. Exercise proper safety practices when using all laboratory equipment, reagents and chemicals. b. Comply with established departmental dress code. 2. Work Practices and Organization a. Adhere to department attendance policies by arrive to lecture/laboratory at the expected time, as denoted in the course syllabus. b. Follow all written instructions. c. Actively listen to verbal instructions. d. Ask quality questions (clarifying, analytical and related to task). e. Submit neat, legible, organized and complete assignments. f. Demonstrate effective time management and complete all tasks within the assignment time frame. g. Keep all laboratory work areas neat, clean and in order. h. Properly care for and use all laboratory equipment. i. Achieve competency and independence in performance of all demonstrated lab skills. 		
	3. Cooperation and Teamwork a. Actively participate in class activities and discussions by: i. Effectively communicating with class members. ii. Showing respect and consideration for other students and		

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	instructors. iii. Willing to share ideas and equally contribute to assigned tasks. b. In laboratory sessions: i. Share resources and equipment. ii. Work cooperatively by adjusting work style and speed. iii. Discuss equitable task allocation and organization prior to performing. 4. Ethics and Professionalism a. Respond maturely to constructive criticism and instruction and make appropriate modifications. b. Seek advice when necessary, admitting limitations when appropriate. c. Recognize and admitting errors. d. Maintain patient confidentiality according to HIPPA regulations. e. Communicate using appropriate terminology and professional procedures. f. Display calm demeanor in all circumstances and maintain work quality under stress.